



Synergistic effects of dietary nano selenium and vitamin C on growth, feeding, and physiological parameters of mahseer fish (*Tor putitora*)



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ABSTRACT

The current study was conducted to determine the synergistic effects of dietary nano selenium (Nano Se) and vitamin C on growth, feeding, and physiological parameters of juvenile mahseer, *Tor putitora*. L-ascorbyl-2-polyphosphate (APP) was used as a source of vitamin C. Four semi-purified experimental diets were prepared. A basal diet kept without the supplementation of any micronutrient and the other three diets were formulated such that three different levels of APP (100, 200, and 300 mg kg⁻¹) were used in combination with a pre-determined dose of Nano Se (0.68 mg kg⁻¹). The results showed that both the micronutrients positively synergized the effects of each other. APP at the rate of 300 mg kg⁻¹ showed strong interaction with Nano Se. The APP₃₀₀ + Nano Se_{0.68} mg kg⁻¹ diet supplemented diet significantly decreased ($P < 0.05$) the feed conversion ratio (FCR) while significantly increased ($P < 0.05$) the weight gain percentage (WG%), feed conversion efficiency (FCE%), specific growth rate (SGR), and serum growth hormone (GH) concentration. Similarly, the physiological parameters such as red blood cells count (RBCs), hemoglobin level (Hb), hematocrit value (Hct), and serum lysozyme activity were also significantly increased in group of fish fed diet supplemented with APP₁₀₀ mg kg⁻¹ in combination with Nano Se_{0.68} mg kg⁻¹ as compared to the control group. The present results clearly indicated the beneficent synergistic effects of Nano Se and APP in mahseer fish. Moreover, the current finding also supported our hypothesis that Nano Se and APP potentiate positively the effect of each other when both the micronutrients are supplemented together in the same fish feed.

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1. Introduction

Minerals and vitamins are two important micronutrients which are required for the maintenance of normal body functions. A proper amount of these micronutrients is required for the normal catalytic processes within the enzymatic system which consists of a variety of enzyme activities linked with the metabolic, endocrine and immune systems (Tomkins, 2002; Keen et al., 2004). The deficiency of these micronutrients causes many metabolic disorders and diseases/infections through their negative influence on the physiological system in fish and other animals (Percival, 1995; Watanabe et al., 1997; Ekiz et al., 2005; Lin and Shiau, 2005). However, their surplus amounts also cause severe toxicity (Watanabe et al., 1997; Hamilton, 2004).

Among micronutrients, selenium and vitamin C both are strong antioxidants and are required for the proper growth and physiological health of animals. Selenium when available in sufficient amount plays an important role in the maintenance of normal body functions of fish and other aquatic species; however, its deficiency weakens the normal physiological health of the body (Ewan, 1976; Raza, 2012; Jamil, 2013; Khan et al., 2016). Similarly, vitamin C is another important micronutrient which is also required in proper amount for the improvement of fish growth and physiological health. This vitamin should be supplemented in fish feed regularly because most of the teleost fish are unable to synthesize ascorbic acid and thus they require proper supplementation of vitamin C in their diet (Al-Amoudi et al., 1992; Dabrowski, 2001; Shalata and Neumann, 2001; Ai et al., 2004; Lin and Shiau, 2005; Khan et al., 2015).

As both micronutrients act as biological antioxidants and help in the protection of cellular membranes from oxidative damage and improve the growth and physiological health of fish and other animals (Ong and Packer, 1992; Wang et al., 1994; Shalata and

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Neumann, 2001; Gaber, 2009; Zuberi et al., 2011; Khan et al., 2015, 2016). Thus due to their ability for interaction, the determination of the synergistic effects among these two important dietary micronutrients may be very useful especially for fish being living in an aquatic environment. By the getting both micronutrients together will enable it to better grow and survive in the current threatening situation of the aquatic environment as currently most water bodies of the world is facing pollution related problems.

T. putitora (Hamilton) belongs to the family Cyprinidae, distributed along the Himalaya belt in Pakistan, Bhutan, and Bangladesh, India, and is an important game and highly valued food fish (Shrestha, 1990; Nautiyal, 1994). Its common name is golden mahseer and it is a column feeder and omnivorous at the adult stage while planktivorous at juvenile stage (Dubey, 1985; Negi, 1994; Shrestha, 1997). Mahseer populations have been declining in most of their natural habitats due to the impacts caused by the industrialization, urbanization, and agricultural developments which are causing ecological alterations and physical changes in the natural environment in lakes and rivers of mid hills (Das and Joshi, 1994; Shrestha, 1994; Joshi, 1988). Another problem is that growth of mahseer in captivity is slower than the growth of fish reared in the wild (Shrestha, 1997). Thus there is a great need to pay attention to the improvement of captive environment for the better and faster growth of this important fish. Some preliminary research studies on the diet development and the use of diets of different nature for feeding mahseer showed beneficent results (Bista et al., 1998). But the existing studies especially about the micronutrients nutrition are not sufficient.

Most of the earlier research work regarding dietary vitamin C and selenium is related to the dietary requirements of these micronutrients separately for various fish species while insufficient literature is available on their combined use in fish feed. In our previous experiments conducted at our laboratory, have already been documented the individual better levels of both of these micronutrients for *T. putitora* (Raza, 2012; Jamil, 2013; Khan et al., 2015, 2016). Thus after determining the individual requirements of these micronutrients and keeping in view their response in the previous studies we made a hypothesis. The hypothesis was that being strong antioxidants, their combined supplementation will synergize the effects of each other and will show better synergistic effects on the growth and physiology of fish as compared to their individual supplementation. The present study was therefore planned to determine the synergistic effects of nano form selenium (Nano Se) and vitamin C (APP) on growth, feeding, and physiological parameters in juvenile mahseer, *T. putitora*.

2. Materials and methods

2.1. Nano Se synthesis

Nano Se was prepared through the precipitation method as described by Jamil (2013) and Khan et al. (2016) for *T. putitora* species and characterized by the X-ray diffraction technique. The calculated average size of the Nano Se was 90 nm.

2.2. Animals and diets

The healthy and uniform sized fish were bought from a fish farm and were transferred to the laboratory. First the fish were acclimatized to the laboratory conditions for about two weeks while feeding it with a control diet. After acclimation, 10 fish (regardless of their sex) of average body weight (2.30 ± 0.04 g) were stocked per tank. The control diet was formulated as shown in Table 1. Four semi-purified and iso-nitrogenous diets consisted of one control diet and three tested diets. The control diet was prepared without

Table 1
Formulation of basal diet for *T. putitora*.

Ingredients	Amount (gram/kilogram)
Soybean meal	130
Wheat bran	10
Fish meal	250
Wheat flour	10
Gluten 60%	500
Sunflower meal	50
Rice bran	10
^a Premix	10
Di-calcium phosphate (DCP)	10
Canola oil	20
Total sum	1,000.00 g kg ⁻¹

^a Premix contained vitamins (except vitamin C), amino acids, and minerals (except selenium).

the supplementation of either test micronutrient while the other three diets were prepared such that three different APP levels (100, 200, and 300 mg APP kg⁻¹ diet) were mixed with a predetermined dose of Nano Se (0.68 mg Nano Se mg kg⁻¹ diet), respectively. All the ingredients were mixed completely in a mixer. Then the control diet (without the mixing of the test micronutrients) and each of the other three diets (each diet contained proper amounts of APP in combination with the predetermined dose of Nano Se) were extruded into pellets. After extrusion, the feed pellets were packed into plastic bags and kept at -20°C until use.

2.3. Experimental design

The 60 day feeding experiment was conducted in a flow-through system. The trial was conducted in a triplicate form and totals 12 fiber tanks were used, with 90 liters of water capacity per tank. Each tank was randomly assigned to one of the three replicates of each of the four dietary treatments. The conditions of the experiment were semi-static and freshwater was used throughout the trial.

The experimental diets were fed to the triplicate groups of fish thrice a day (6:00a.m.; 12:00p.m.; 5:00p.m.) until apparent satiation. The water quality parameters were maintained at the proper levels as reported for the under study species by Khan et al. (2015). During the experimental period, the average water pH ranged from 7 to 8 and temperature ranged from 21 to 23 °C. The ammonia level measured was lower than 0.20 mg L⁻¹, while the dissolve oxygen content (DO₂) ranged from 6 to 6.5 mg L⁻¹.

2.4. Samples collection

At the end of the 60 day experiment, fish were fasted for 24 h prior to sampling to allow the evacuation of food in the gastrointestinal tract. On the following day, fish were sampled as follows; One by one all fish were removed from each tank and euthanized in a tricaine methanesulfonate solution (MS 222, 0.5 g L⁻¹; Argent Chemical Laboratories, Redmount, WA, USA) respectively. Each euthanized fish was blotted dry on the paper towels and weighed. After weighting, all fish per tank were subjected to the blood collection process and the blood was collected by the tail ablation method (Khan et al., 2015, 2016). The remaining blood was centrifuged at 3500 rpm for 15 min. The serum was separated and stored at -20°C for the analysis of the serum growth hormone concentration and serum lysozyme activity.

2.5. Growth and feeding parameters

The weight gain percentage (WG%), feed conversion ratio (FCR), feed conversion efficiency (FCE%), and specific growth rate (SGR%), was calculated according to the method followed by Khan et al. (2015).

Table 2

Synergistic effects of the diets supplemented with different levels of APP in combination with Nano Se on the growth and feeding parameters of juvenile *T. putitora*.

Parameters/Diets	WG%	SGR (%)	FCE (%)	FCR
Basal diet	64.6 ± 0.52 ^g	0.40 ± 0.02 ^g	28.06 ± 0.13 ^g	3.56 ± 0.06 ^a
APP ₁₀₀ + Nano Se _{0.68}	98.76 ± 1.76 ^d	0.59 ± 0.09 ^d	42.43 ± 0.36 ^d	2.35 ± 0.01 ^d
APP ₂₀₀ + Nano Se _{0.68}	104.46 ± 0.70 ^c	0.62 ± 0.01 ^c	45.24 ± 0.11 ^c	2.21 ± 0.04 ^e
APP ₃₀₀ + Nano Se _{0.68}	125.41 ± 0.74 ^a	0.72 ± 0.03 ^a	50.93 ± 0.08 ^a	1.96 ± 0.04 ^g

Data are represented as Mean ± SE (n = 3). Means followed by a different letter within a column are significantly different (P < 0.05) (ANOVA followed by Tukey's LSD test).

2.6. Serum growth hormone assay

For the analysis of serum growth hormone, the method described by Khan et al. (2015, 2016) was followed. Serum GH concentration in all group units was estimated by using a Micro ELISA HGH kit (Amgenix MicroLISATM, USA).

2.7. Physiological parameters

About 1 ml blood was collected in VACUETTE EDTA K3 tubes for the analysis of hematological parameters while the rest of blood was centrifuged and was used for the analysis of serum lysozyme activity. The blood parameters such as, red blood cell (RBC) count (106 μL^{-1}), hemoglobin (Hb) level (g dL⁻¹), and hematocrit (Hct%) were determined by using the hematology analyzer (Sysmax KX-21 Japan).

An important innate immunity parameter, the lysozyme activity was determined according to the method followed by Khan et al. (2016). The absorbance was observed with the help of spectrophotometer at 450 nm after 1 min intervals for 10 min and the lysozyme activity ($\mu\text{g mL}^{-1}$) was calculated by using hen egg white lysozyme (Sigma-Aldrich) as a standard.

2.8. Statistical analysis

Obtained results were analyzed through one-way analysis of variance (ANOVA) by using the Statistix 8.1 analytical software. When any significant difference was detected, a Tukey's LSD (least significant difference) test was used to compare the means. Treatment differences were measured with the significance level at (P < 0.05).

3. Results

3.1. Growth and feeding parameters

No mortality was observed. The synergistic effect of dietary Nano Se along with different levels of APP on the growth and feeding parameters of *T. putitora* is shown in Table 2. The addition of dietary Nano Se in combination with APP into the fish feed showed significant effect on the growth and feeding parameters of *T. putitora*. The dietary Nano Se and APP supplementation

synergized the effect of each other and significantly higher (P < 0.05) weight gain percentage (WG%), feed conversion efficiency (FCE%), specific growth rate (SGR%), while significantly lower (P < 0.05) feed conversion ratio (FCR) was observed in the group of fish fed APP₃₀₀ + Nano Se_{0.68} mg kg⁻¹.

3.2. Serum growth hormone level

Serum growth hormone results are given in Table 3. The lowest GH level was observed in group of fish reared on the basal diet. However, the supplementation of dietary Nano Se along with APP significantly increased the serum growth hormone concentration

Table 3

Synergistic effects of the diets supplemented with different levels of APP in combination with Nano Se on the serum growth hormone level in juvenile *T. putitora*.

Parameters/diets	Serum growth hormone (ng mL ⁻¹)
Basal diet	0.09 ± 0.06 ^a
APP ₁₀₀ + Nano Se _{0.68}	0.100 ± 0.08 ^b
APP ₂₀₀ + Nano Se _{0.68}	0.115 ± 0.06 ^c
APP ₃₀₀ + Nano Se _{0.68}	0.312 ± 0.09 ^d

Data are represented as Mean ± SE (n = 3). Means are followed by a different letter within a column are significantly different (P < 0.05) (ANOVA followed by Tukey's LSD test).

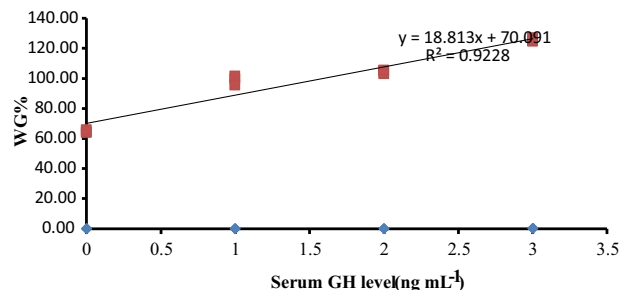


Fig. 1. Positive co-relation between the WG% and serum growth hormone level.

Table 4

Synergistic effects of the diets supplemented with different levels of APP in combination with Nano Se on the hematology of juvenile *T. putitora*.

Parameters/ diets	RBC count (106 μL)	Hb level (g dL ⁻¹)	Hct (%)
Basal diet	2.26 ± 0.08 ^a	7.3 ± 0.20 ^a	32.92 ± 0.01 ^a
APP ₁₀₀ + Nano Se _{0.68}	2.64 ± 0.13 ^b	8 ± 0.05 ^c	33.96 ± 0.03 ^b
APP ₂₀₀ + Nano Se _{0.68}	2.98 ± 0.03 ^c	8.4 ± 0.07 ^c	34.53 ± 0.08 ^c
APP ₃₀₀ + Nano Se _{0.68}	3.2 ± 0.05 ^d	9.16 ± 0.14 ^d	36.4 ± 0.30 ^d

Data are represented as Mean ± SE (n = 3). Means are followed by a different letter within a column are significantly different (P < 0.05) (ANOVA followed by Tukey's LSD test).

in *T. putitora*. The supplementation of APP at the rate of 300 mg kg⁻¹ in combination with Nano Se at the rate of 0.68 mg kg⁻¹ showed strong synergistic effects and significantly increased (P < 0.05) the serum growth hormone concentration as compared to the diets contained lower doses of APP in combination with Nano Se as well the control diet.

Moreover, a positive linear co-relation between the weight gain percentage (WG%) and the serum growth hormone level of *T. putitora* which is shown in Fig. 1.

3.3. Physiological parameters

The effects of dietary Nano Se in combination with different levels of APP on the hematological parameters like red blood cells (RBCs) count, hemoglobin (Hb) level, and hematocrit (Hct%) are

shown in Table 4. The dietary Se Nano supplementation in combination with APP showed positive synergistic effects on the hematology of *T. putitora*. Thus, the hematological parameters like RBCs count, Hb level, and Hct%, were significantly increased (P < 0.05) in the group of fish fed diet supplemented with dietary APP₃₀₀ mg kg⁻¹ in combination with the dietary Nano Se_{0.68} mg kg⁻¹ as compared to the control group.

Similarly, the serum lysozyme activity was also significantly increased with APP₃₀₀ + Nano Se_{0.68} mg kg⁻¹ supplementation as compared to the control diet. The serum lysozyme activity results are shown in Table 5.

Table 5

Synergistic effects of the diets supplemented with different levels of APP in combination with Nano Se on the serum lysozyme activity of juvenile *T. putitora*.

Parameters/Diets	Serum lysozyme activity ($\mu\text{mol min}^{-1} \text{mg}^{-1} \text{protein}$)
Basal diet	0.88 \pm 0.08 ^a
APP ₁₀₀ + Nano Se _{0.68}	2.40 \pm 0.06 ^b
APP ₂₀₀ + Nano Se _{0.68}	3.28 \pm 0.07 ^c
APP ₃₀₀ + Nano Se _{0.68}	4.31 \pm 0.06 ^d

Data are represented as Mean \pm SE, (n = 3). Means are followed by different letter within the column are significantly different (P < 0.05). (ANOVA followed by Tukey's LSD test).

4. Discussion

Initial studies from our laboratory have already been determined the better dietary levels of both nano selenium (Nano Se) 0.68 mg kg⁻¹ (Jamil, 2013; Khan et al., 2016), and vitamin C, l-ascorbyl-2-polyphosphate (APP) 300 mg kg⁻¹ diet (Khan et al., 2015) respectively, for *T. putitora*. As some of the earlier research studies showed that synergistic interactions exist between the selenium and other vitamins such as E (Gatlin et al., 1986; Fonseca et al., 2013). However, no information is available about the synergistic effects of dietary vitamin C and selenium in fish. Thus the present study was designed on the basis of the hypothesis that the dietary Nano Se when use in combination with APP in same fish feed, may show strong synergistic interactions. The results obtained revealed that these two important micronutrients when supplied in combination showed strong synergistic interactions and positively potentiated the growth, feeding, and physiological parameters of *T. putitora*.

Earlier research studies showed that adequate amount of dietary selenium is essential for the proper body growth of fish and other animals (Hoffmann and Berry, 2008; Schrauzer and Surai, 2009; Ewan, 1976; Raza, 2012; Khan et al., 2016; Jamil, 2013). In a previous study, it was observed that the diet enriched with vitamin C, E and Se significantly increased the weight gain in tilapia (*O. niloticus*) (Kim and Mahan, 2003). Selenium supplementation also increases the selenium concentration in the muscle tissue of some fish species (Buckley, 2000). Fonseca et al. (2013) observed that the tilapia fillet fed the diet supplemented with 400 mg kg⁻¹ vitamin C and vitamin E, respectively and 0.4 mg kg⁻¹ selenium oxide showed significantly increased (P < 0.05) weight gain (WG) and feed conversion efficiency (FCE%) as compared to the control diet. The higher growth of tilapia fed the increasing levels of selenium, vitamins C and E can be explained by the facts that both selenium and vitamin C are important micronutrients which are needed for the better growth performance and feeding parameters of fish (NRC, 1993; Wang et al., 2003; Khan et al., 2015, 2016). Their individual supplementation enhances erythrocytes production and prevents high quantity fat accumulation in the liver tissue (Martins et al., 2008; Khan et al., 2015, 2016). The present study also revealed a strong synergistic interaction among dietary Nano Se and APP. Both the micronutrients when supplemented in combination in *T. putitora* feed resulted in significantly increased growth and feeding parameters (weight gain percentage, feed conversion efficiency, feed conversion ratio, specific growth ratio) and significantly decreased the feed conversion ratio.

Like selenium, vitamin C is another essential micronutrient which improves the growth and physiological health of fish. Beside a strong antioxidant, it also accelerates the growth hormone production and thus indirectly increases the body growth in fish (Lovell, 1989; Eipper et al., 1993; James and Manju, 2001; Wang et al., 2003; Kumari and Sahoo, 2005; Ai et al., 2004; Ai et al., 2006; Affonso et al., 2007; Aysun, 2009; Jimenez-Fernandez et al., 2012; Denny-Brown et al., 2012; Zhou et al., 2012; Ullah, 2012;

Moein, 2012; Khan et al., 2015; Tran et al., 2006). In the present study, we observed that the dietary APP and Nano Se when used in combination in the same fish feed, strongly synergized the effects of each other. The growth performance and feeding parameters (WG%, SGR%, FCR, FCE%, and serum growth hormone levels) of *T. putitora* were significantly increased by consuming the same feed which was supplemented with the dietary Nano Se (at the rate of 0.68 mg kg⁻¹ diet) and APP (at the rate of 300 mg kg⁻¹). These results were more profound than those observed in our previous experiments (Khan et al., 2015, 2016).

The strong antioxidant property of both the selenium (Ong and Packer, 1992; El-Hammady et al., 2007; Molnár et al., 2011; Khan et al., 2016) and vitamin C (Waagbo et al., 1993; Soliman et al., 1994; Chen et al., 2003; Sahoo and Mukherjee, 2003; Nayak et al., 2007; Affonso et al., 2007; NRC, 2011; Pimpimol et al., 2012; Khan et al., 2015) might provide stability and integrity to the blood cells of fish and thus protect them from hemolysis during the normal metabolism and stressful conditions. In the present finding, when both the micronutrients were supplemented together in the same fish feed, it documented that the APP₃₀₀ + Nano Se_{0.68} supplemented diet significantly improved the hematological parameters (RBCs count, Hb level, and Hct%) in juvenile *T. putitora* as compared to the control diet and the diets that contained the low levels of APP in combination with Nano Se. These synergistic effects were significantly better than those observed with the individual supplementation of either vitamin C or selenium in our previous experiments (Khan et al., 2015, 2016).

In fish, the non-specific immune system as compared to the specific immune system is playing a significant role against stress induced by diseases, pollution, and other stressful objects (Anderson, 1992). The supplementation of selenium (Burk et al., 2003; Kumar et al., 2008; Lee et al., 2009; Choi et al., 2013; Khan et al., 2015) and vitamin C (Guha et al., 1993; Fetoui et al., 2008; Korkmaz et al., 2009; Khan, 2014) improves the production of B-lymphocytes. As B-lymphocytes production plays a significant role in the improvement of the lysozyme activity of fish. Thus the improved lysozyme activity then enhances the immunity of fish.

Currently, we supplemented APP and Nano Se in combination in the same diet for observing their synergistic effects. The APP at the rate of 300 mg kg⁻¹ showed strong synergistic effects with Nano Se_{0.68} mg kg⁻¹ and significantly increased the serum lysozyme activity as compared to the previous feeding trials regarding the micronutrients requirement determination for *T. putitora* (Khan et al., 2015, 2016).

Furthermore, a positive co-relation was observed in the present study among the serum GH concentration and the WG% in *T. putitora* in response to the diets supplemented with dietary Nano Se along with different levels of APP. As in several teleost fish, higher circulating GH levels result in increased feed conversion efficiency and growth rates (Collipp et al., 1984; Won and Borski, 2013; Khan et al., 2015, 2016). The positive co-relation among the serum GH level and the WG% of this study was stronger as compared to the co-relation which was observed in our previous experiments (Khan et al., 2015, 2016).

In conclusion, the results of the present study indicated that dietary vitamin C (at the rate of APP₃₀₀ mg kg⁻¹ diet) showed strong synergistic interaction along with Nano Se_{0.68} mg kg⁻¹ diet. The APP₃₀₀ + Nano Se_{0.68} mg kg⁻¹ supplemented diet significantly increased the growth, feeding, and physiological parameters of *T. putitora*. The results of the present study are significantly better than those obtained in our previous experiments about the vitamin C and Nano Se individual requirements for the same fish species. Thus, this study supported our hypothesis and furthermore, it recommends the supplementation of APP₃₀₀ + Se Nano_{0.68} mg kg⁻¹ in

juvenile *T. putitora* feed for better growth, feeding, and physiological parameters.

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References

- Affonso, E.G., Silva, E., Tavares-Dias, M., de Menezes, G., de Carvalho, C., Nunes, É., Ituassú, D., Roubach, R., Ono, E., Fim, J., Marcon, J., 2007. Effect of high levels of dietary vitamin C on the blood responses of matrinxã (*Brycon amazonicus*). *Comp. Biochem. Physiol.* 147, 383–388.
- Ai, Q., Mai, K., Zhang, C., Xu, W., Duan, Q., Tan, B., Liufu, Z., 2004. Effects of dietary vitamin C on growth and immune response of Japanese seabass (*Lateolabrax japonicus*). *Aquaculture* 242, 489–500.
- Ai, Q., Mai, K., Tan, B., Xu, W., Zhang, W., Ma, H., Liufu, Z., 2006. Effects of dietary vitamin C on survival, growth, and immunity of large yellow croaker (*Pseudosciaena crocea*). *Aquaculture* 261, 327–336.
- Al-Amoudi, M.M., El-Nakkadi, A.M.N., El-Nouman, B.M., 1992. Evaluation of optimum dietary requirement of vitamin C for the growth of *Oreochromis spilurus* fingerlings in water from the Red Sea. *Aquaculture* 105, 165–173.
- Anderson, D.P., 1992. Immuno-stimulants, adjuvants, and vaccine carriers in fish: application to aquaculture. *Annu. Rev. Fish Dis.* 2, 281–307.
- Aysun, H., 2009. An Overview of ascorbic acid biochemistry. *J. Fac. Pharm. Ankara (Turkey)* 38, 233–255.
- Bista, J.D., Shrestha, R.K., Yamada, O., 1998. On the relationship of GSI, total lipid and moisture content of mahseer (*Tor putitora*). In: Pradhan, B.R., Wagle, S.K., Osamu, Y., Takano (Eds.), *Present Status of Fisheries Research, Development and Education*. pp. 68–70.
- Buckley, W.T., 2000. Trace element dynamics. In: Mello, D., J.P.F. (Eds.), *Farm Animal Metabolism and Nutrition*. CAB International, Wallingford, UK, pp. 161–182.
- Burk, R.F., Hill, K.E., Motley, A.K., 2003. Selenoprotein metabolism and function: evidence for more than one function for selenoprotein. *J. Nutr.* 133, 1517–1520.
- Chen, R., Lochmann, R., Goodwin, A., Praveen, K., Dabrowski, K., Lee, K.J., 2003. Alternative complement activity and resistance to heat stress in golden shiners (*Notemigonus crysoleucas*) are increased by dietary vitamin C levels in excess of requirements for prevention of deficiency signs. *J. Nutr.* 133, 2281–2286.
- Choi, Y.J., Kim, N.N., Shin, H.S., Park, M.S., Kil, G.S., Choi, C.Y., 2013. Effects of waterborne selenium exposure on the antioxidant and immunological activity in the goldfish (*Carassius auratus*). *Mol. Cell. Toxicol.* 9, 365–373.
- Collipp, P.J., Kelemen, J., Chen, S.Y., Castro-magana, M., Angulo, M., Derenoncourt, A., 1984. Growth hormone inhibition causes increased selenium levels in Duchenne muscular dystrophy: a possible new approach to therapy. *J. Med. Genet.* 21, 254–256.
- Dabrowski, K., 2001. History, present, and future of ascorbic acid research in aquatic organisms. In: Dabrowski, K. (Ed.), *Ascorbic Acid in Aquatic Organisms-status and Perspectives*. CRC Press, Boca Raton, pp. 255–277.
- Das, P., Joshi, K.D., 1994. Mahseer conservation-present and future. Mahseer; the game fish, D3-D9. Nautiyal, P., (Comp. and Ed.), Rachna Publisher, Garhwal, U.P., India.
- Denny-Brown, S., Stanley, T.L., Grinspoon, S.K., Makimura, H., 2012. The association of macro- and micronutrient intake with growth hormone secretion. *Growth Horm. IGF Res.* 22, 102–107.
- Dubey, G.P., 1985. Conservation of dying king mahseer the mighty game fish and its future role in reservoir fisheries. *Fish. Bull. (Punjab, India)* 9, 182.
- Eipper, B.A., Milgram, S.L., Husten, E.J., Yun, H.Y., Mains, R.E., 1993. Peptidylglycine alpha-amidating monooxygenase: multifunctional protein with catalytic, processing, and routing domains. *Protein Sci.* 2, 489–497.
- Ekiz, C., Agaoglu, L., Karakas, Z., Gurel, N., Yalcin, I., 2005. The effect of iron deficiency anemia on the function of the immune system. *Hematol. J.* 5, 579–583.
- El-Hammady, A.K.I., Ibrahim, S.A., El-Kasheif, M.A., 2007. Synergistic reactions between vitamin E and Selenium in diets of hybrid tilapia (*Oreochromis niloticus* × *Oreochromis aureus*) and their effect on the growth and liver histological structure. *Egypt. J. Aquat. Biol. Fish.* 1, 53–58.
- Ewan, R.C., 1976. Effect of selenium on rat growth, growth hormone and diet utilization. *J. Nutr.* 106, 702–709.
- Fetoui, H., Garoui, E.M., Makni-ayadi, F., Zeghal, N., 2008. Oxidative stress induced by lambda-cyhalothrin (LTC) in rat erythrocytes and brain: attenuation by vitamin C. *Environ. Toxicol. Pharmacol.* 26, 225–231.
- Fonseca, S.B.D., Silva, J.H.V.D., Mendes, E.M.B., Fernandes, J.B.K., Amancio, A.L.L., Filho, J.J., Lacerda, P.B.D., Silva, F.R.P.D., 2013. Influence of levels and forms of selenium associated with levels of vitamins C and E on the performance, yield and composition of tilapia fillet. *Food Sci. Technol. (Campinas, Brazil)* 33, 109–115.
- Gaber, M.M., 2009. Efficiency of selenium ion inclusion into common carp (*Cyprinus carpio* L.) diets. *Afr. J. Agric. Res.* 4, 348–353.
- Gatlin III, D.M., Poe, W.E., Wilson, R.P., 1986. Effects of singular and combined dietary deficiencies of selenium and vitamin E on fingerling catfish. *J. Nutr.* 116, 1061–1067.
- Guha, D., Dutta, K., Das, M., 1993. Vitamin C as antitoxic factor in DDT induced haematotoxicity in *Clarias batrachus*. *Proc. Zool. Soc. (Calcutta, India)* 46, 11–15.
- Hamilton, S.J., 2004. Review of selenium toxicity in the aquatic food chain. *Sci. Total Environ.* 326, 1–31.
- Hoffmann, P.R., Berry, M.J., 2008. The influence of selenium on immune responses. *Mol. Nutr. Food Res.* 52, 1273–1280.
- James, D.C., Manju, B.R., 2001. Effect of ascorbic acid intake on nonheme-iron absorption from a complete diet 1, 2. *Am. J. Clin. Nutr.* 73, 93–98.
- Jamil, Z., 2013. Effects of Inorganic and Nanoform of Selenium on Growth Performance and Biochemical Indices of Mahseer (*Tor Putitora*). MPhil. Thesis. Department of Animal Sciences, Faculty of Biological Sciences, Islamabad, Pakistan.
- Jimenez-Fernandez, E., Ponce, M., Zuasti, E., Fernandez-Diaz, C., Manchado, M., Infante, C., 2012. Molecular characterization and transcriptional regulation of the sodium-dependent vitamin C transporter genes (slc23a1 and slc23a2) in a teleost fish, the Senegalese sole (*Solea senegalensis*). *Comp. Biochem. Physiol. (Part B)* 161, 208–218.
- Joshi, C.B., 1988. Mahseer fishery of some hill streams in western Himalayas. *Indian J. Fish.* 35, 327–329.
- Keen, C.L., Uriu-Adams, J.Y., Emsimsa, J.L., Gershwin, M.E., 2004. Trace elements/minerals and immunity. In: Gershwin, M.E., Nestel, P., Keen, C.L., Totowa, N.J. (Eds.), *Handbook of Nutrition and Immunity*. Humana Press, pp. 117–140.
- Khan, K.U., 2014. Synergistic Effects of Vitamin C and Selenium (Nano Form) on Growth, Biochemical and Immunological Indices of Juvenile Mahseer (*Tor Putitora*). MPhil. Thesis. Department of Animal Sciences, Faculty of Biological Sciences, Quaid-i-Azam University, Islamabad, Pakistan (Thesis&).
- Khan, K.U., Zuberi, A., Ullah, I., Sajjad, 2015. Effects of graded level of dietary L-ascorbyl-2-polyphosphate on growth performance and some hematological indices of juvenile mahseer (*Tor putitora*). *Int. J. Agric. Biol.* 17, 821–827.
- Khan, K.U., Zuberi, A., Nazir, S., Fernandes, J.B.K., Jamil, Z., Sarwar, H., 2016. Effects of dietary selenium nanoparticles on physiological and biochemical aspects of juvenile *Tor putitora*. *Turk. J. Zool.* 40, 704–712.
- Kim, Y.Y., Mahan, D.C., 2003. Biological aspects of selenium in farm animals. *Asian-Australas. J. Anim. Sci.* 16, 435–444.
- Korkmaz, N., Cengiz, E.I., Unlu, E., Uysal, E., Yanar, M., 2009. Cypermethrin-induced histopathological and biochemical changes in Nile tilapia (*Oreochromis niloticus*) and the protective and recuperative effect of ascorbic acid. *Environ. Toxicol. Pharmacol.* 28, 198–205.
- Kumar, N., Garg, A.K., Mudgal, V., 2008. Effect of different levels of selenium supplementation on growth rate, nutrient utilization, blood metabolic profile, and immune response in lambs. *Biol. Trace Elem. Res.* 126, S44–S46.
- Kumari, J., Sahoo, P.K., 2005. High dietary vitamin C affects growth, non-specific immune responses and disease resistance in Asian catfish (*Clarias batrachus*). *Mol. Cell. Biochem.* 280, 25–33.
- Lee, J.H., Kim, Y.C., Park, S.L., Bai, S.C., 2009. Evaluation of the optimum dietary selenium (Se) level to improve immune responses in juvenile olive flounder (*Paralichthys olivaceus*). *J. Korean Fish. Soci.* 42, 26–33.
- Lin, Y., Shiau, S., 2005. Dietary selenium requirements of juvenile grouper (*Epinephelus malabaricus*). *Aquaculture* 250, 356–363.
- Lovell, R.T., 1989. Vitamin C (ascorbic acid). In: *Nutrition and Feeding of Fish. An AVI Book. Van Nostrand Reinhold Publication*, pp. 54–60.
- Martins, M.L., Miyazaki, D.M.Y., Moraes, F.R.D., Ghiraldelli, L., Adamante, W.D.B., Mourão, J.L.P., 2008. Ration supplemented with vitamin C-E influences the acute inflammatory response in Nile tilapia. *Ciência Rural (Santa Maria, Brazil)* 38, 213–218.
- Moein, F., 2012. Effect of dietary vitamin C on growth and feeding parameters, carcass composition and survival rate of common carp (*Cyprinus carpio*). *Glob. Vet.* 8, 507–510.
- Molnár, T., Biró, J., Balogh, K., Mézes, M., Hancz, C., 2011. Improving the nutritional value of Nile tilapia fillet by dietary selenium supplementation. *Israel J. Aquacult.-Bamidgheh* 64, 1–6.
- National research council (NRC), 1993. *Nutrient Requirements of Fish*. National Academy Press, Washington DC, USA, p. 114.
- National Research Council (NRC), 2011. *Nutrient Requirements of Fish and Prawn*. National Academies Press, Washington, DC, USA, pp. 207–209.
- Nautiyal, P., 1994. The endangered himalayan mahseer. a decade of retrospection. In: Dehadrai, P.V., Das, P., Verma, S.R. (Eds.), *Threatened Fishes of India. Proceedings of The National Seminar on Endangered Fishes of India Held at National Bureau of Fish Genetic Resources, Allahabad, 25–26 April, 1992*. NATCON Publication, Muzaffarnagar, pp. 191–196.
- Nayak, S.K., Swain, P., Mukherjee, S.C., 2007. Effect of dietary supplementation of probiotic and vitamin C on the immune response of Indian major carp (*Labeo rohita*) (Himalaya). *Fish Shellfish Immunol.* 23, 892–896.
- Negi, S.S., 1994. *Himalaya Fishes and Fisheries*. Ashish Publishing, New Delhi, India, pp. 291.
- Ong, A.S.H., Packer, L., 1992. Lipid-Soluble antioxidants: biochemistry and clinical applications. *Mol. Cell. Biol. Updates (Basel, Birkhäuser)*, 642.
- Percival, S.S., 1995. Neuropenia caused by copper deficiency: possible mechanisms of action. *Nutr. Rev.* 53, 59–66.

- Pimpimol, T., Phoosamran, K., Chitmanat, C., 2012. Effect of dietary vitamin C supplementation on the blood parameters of mekong giant catfish (*Pangasianodon gigas*). *Int. J. Agric. Biol.* 14, 256–260.
- Raza, A., 2012. Effects of Graded Levels of Dietary Selenium Supplementation on The Growth of Juvenile Mahseer (*Tor Putitora*). MPhil. Thesis. Department of Animal Sciences, Faculty of Biological Sciences, Islamabad, Pakistan.
- Sahoo, P.K., Mukherjee, S.C., 2003. Immunomodulation by dietary vitamin C in healthy and aflatoxin B1-induced immune compromised rohu (*Labeo rohita*). *Comp. Immunol. Microbiol. Infect. Dis.* 26, 65–76.
- Shalata, A., Neumann, P.M., 2001. Exogenous ascorbic acid (vitamin C) increases resistance to salt stress and reduces lipid peroxidation. *J. Exp. Bot.* 52, 2207–2211.
- Shrestha, T.K., 1990. Behaviour of golden mahseer, *T. putitora* (Ham) in nature and captivity. *J. Freshw. Biol.* 23, 209–210.
- Shrestha, T.K., 1994. Development of mahseer culture towards ranching. Nautiyal, P. (Comp. and Ed.), Mahseer; The Game Fish, D26-D41. Rachna, Srinagar, India.
- Shrestha, T.K., 1997. Prospects of propagating the mahseer in Phewa Lake of the Pokhara Valley, India. Mahseer, 70–71.
- Soliman, A.K., Jauncey, K., Roberts, R.J., 1994. Water-soluble vitamin requirements of tilapia: ascorbic acid (vitamin C) requirement of Nile tilapia (*Oreochromis niloticus* L.). *Aquacult. Res.* 25, 269–278.
- Tomkins, A.M., 2002. Nutrition, infection and immunity: public health implications. In: Calder, P.C., Field, C.J., Gill, H.S. (Eds.), *Nutrition and Immune Function*. *Frontiers in Nutrition Science*. No.1. CABI Publishing, Wallingford, UK, pp. 375–412.
- Tran, C.D., Diorio, C., Berube, S., 2006. Relation of insulin-like growth factor (IGF) I and IGF-binding protein 3 concentrations with intakes of fruit, vegetables, and antioxidants. *Am. J. Clin. Nutr.* 84, 1518–1526.
- Ullah, N., 2012. Effect of Different Levels of Dietary Ascorbic Acids on Growth, Liver Vitamin C and Response to Hypoxic and Hyperoxic Stress in Juvenile Silver Carp (*Hypophthalmichthys Molitrix*). MPhil. Thesis. Department of Animal Sciences, Faculty of Biological Sciences, Islamabad, Pakistan.
- Waagbo, R., Glette, J., Raa-Nilsen, E., Sandnes, K., 1993. Dietary vitamin C, immunity, and disease resistance in Atlantic salmon (*Salmo salar*). *Fish Physiol. Biochem.* 12, 61–73.
- Wang, A.I., Wang, W.N., Liu, C.Q., Wang, S.A., 1994. Effects of selenium concentrations in feed on the growth and selenium contents of *Penaeus chinensis*. *J. Fish. (China)* 18, 245–248.
- Wang, X., Kim, K., Bai, S.C., Huh, M., Cho, B., 2003. Effects of the different levels of dietary vitamin C on growth and tissue ascorbic acid changes in parrot fish (*Oplegnathus fasciatus*). *Aquaculture* 215, 203–211.
- Watanabe, T., Kiron, V., Satoh, S., 1997. Trace minerals in fish nutrition. *Aquaculture* 151, 185–207.
- Won, E.T., Borski, R.J., 2013. Endocrine regulation of compensatory growth in fish. *Front. Endocrinol.* 4, 1–13.
- Zhou, Q., Wang, L., Wang, H., Xie, F., Wang, T., 2012. Effect of dietary vitamin C on the growth performance and innate immunity of juvenile cobia (*Rachycentron canadum*). *Fish Shellfish Immunol.* 32, 969–975.
- Zuberi, A., Gima, M., Gima, A., Huston, A., Chaimongkol, A., Li, M., Umali-Maceina, G., Dunham, R., 2011. Diet fed to brood stock effects the growth of channel catfish (*Ictalurus punctatus* L.) fry. *Aquacult. Res.* 42, 1899–1904.